The FASEB Journal www.fasebj.org

Published online before print July 19, 2016, doi: 10.1096/fj.201600646R November 2016 <u>The FASEB Journal</u> vol. 30 no. 11 3682–3689

Cannabinoids, inflammation, and fibrosis

Robert B. Zurier¹ and Sumner H. Burstein

+ Author Affiliations

→¹Correspondence: Department of Biochemistry and Molecular Pharmacology, University of Massachusetts Medical School, 364 Plantation St., Worcester, MA 01605, USA. E-mail: robert.zurier@umassmed.edu

Abstract

Cannabinoids apparently act on inflammation through mechanisms different from those of agents such as nonsteroidal anti-inflammatory drugs (NSAIDs). As a class, the cannabinoids are generally free from the adverse effects associated with NSAIDs. Their clinical development thus provides a new approach to treatment of diseases characterized by acute and chronic inflammation and fibrosis. A concise survey of the anti-inflammatory actions of the phytocannabinoids Δ^{9} tetrahydrocannabinol (THC), cannabidiol, cannabichromene, and cannabinol is presented. Mention is also made of the noncannabinoid plant components and pyrolysis products, followed by a discussion of 3 synthetic preparations-Cesamet (nabilone; Meda Pharmaceuticals, Somerset, NJ, USA), Marinol (dronabinol; THC; AbbVie, Inc., North Chicago, IL, USA), and Sativex (Cannabis extract; GW Pharmaceuticals, Cambridge United Kingdom)—that have anti-inflammatory effects. A fourth synthetic cannabinoid, ajulemic acid (AJA; CT-3; Resunab; Corbus Pharmaceuticals, Norwood, MA, USA), is discussed in greater detail because it represents the most recent advance in this area and is currently undergoing 3 phase 2 clinical trials by Corbus Pharmaceuticals. The endogenous cannabinoids, including the closely related lipoamino acids, are then discussed. The review concludes with a presentation of a possible mechanism for the anti-inflammatory and antifibrotic actions of these substances. Thus, several cannabinoids may be considered candidates for development as anti-inflammatory and antifibrotic agents. Of special interest is their possible use for treatment of chronic inflammation, a major unmet medical need.-Zurier, R. B., Burstein, S. H. Cannabinoids, inflammation, and fibrosis.

endocannabinoids specialized proresolving mediators anti-inflammatory antifibrotic

Preparations derived from Cannabis have been the source of medical therapies since the earliest records on pharmacobotany (1). Many beneficial effects of Cannabis on the human body, including those on "rheumatism" were noted 4000 yr ago in a work reported by Hui-Lin Li called *Pen-tsao* (2). The term cannabinoid usually refers to compounds that activate the G-protein-coupled cannabinoid receptors 1 and 2 (CB1 and -2). CB1 receptors, located mainly on neurons in the hippocampus and basal ganglia, mediate the psychoactive actions of cannabinoids (3). CB2 receptors are present mainly on tissue and circulating cells of the immune system (4). However, many Cannabis components that do not activate either receptor are sometimes called cannabinoids. Given that the Cannabis plant contains more than 60 cannabinoids and 200-250 noncannabinoid constituents, it follows that the therapeutic benefits of marijuana are related to some combination of these compounds. We review the current knowledge of the mechanisms whereby phytocannabinoids, noncannabinoid plant components, and their pyrolysis products aid in the control of inflammation and fibrosis. We also address the development of synthetic cannabinoids as treatment for patients with diseases characterized by chronic inflammation and subsequent fibrosis. The ability of some cannabinoids to facilitate the resolution of inflammation by stimulating the action of several specialized proresolving mediators (SPMs), an important emerging concept, is also discussed. The roles of endogenous cannabinoids (endocannabinoids) and the closely related lipoamino acids in control of inflammation are also discussed.

CANNABIS CONSTITUENTS

Experiments with Δ^9 -tetrahydrocannabinol (THC; Fig.1), the main psychoactive cannabinoid in the plant, have been helpful in understanding the antiinflammatory actions of the nonpsychoactive cannabinoids (5, 6). Although there is rich documentation of the anti-inflammatory actions of several of the nonpsychoactive constituents, that literature has been crowded out by discussions about psychoactivity and legalization of marijuana. For example, the noncannabinoid, prenylated flavone cannflavin is 30 times more potent as an inhibitor of cyclooxygenase (COX) than the time-honored anti-inflammatory drug aspirin. In addition, Sofia et al. (7, 8) demonstrated the anti-inflammatory actions of a crude extract of THC and of the nonpsychoactive Cannabis constituents cannabidiol (CBD) and cannabinol in a carrageenan-induced paw edema model of acute inflammation in rats. They showed in the same model that THC is 80 times more potent than aspirin and twice as potent as hydrocortisone and that the nonpsychoactive constituent cannabichromene also suppresses the induced inflammation (94-11). Moreover, CBD reduced acute inflammation in a murine model of collagen-induced arthritis (12). The precise mechanisms whereby CBD reduces inflammation are not clear. CBD does reduce production of the proinflammatory cytokine TNF- α and induces reduction of fatty acid aminohydrolase (FAAH) activity, thereby increasing production of anandamide, an anti-inflammatory endocannabinoid. Volatile oil components of Cannabis sativa suppress COX1 activity (13, 14), and pyrolysis products of CBD exhibit activity in COX-1-suppression assays (15). Several of the most abundant cannabinoid and noncannabinoid constituents of the plant are not psychoactive (16). Thus, it is clear that cannabinoid and noncannabinoid constituents of Cannabis are potential nonpsychoactive anti-inflammatory agents.

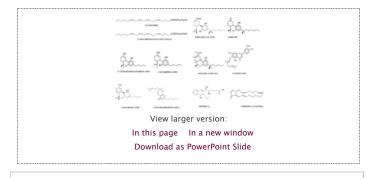


Figure 1.

Structures of compounds discussed in this review.

As the most abundant nonpsychoactive cannabinoid in *Cannabis*, CBD has been studied extensively for its anti-inflammatory properties. As noted, it is active in a murine model of collagen-induced arthritis. In addition, CBD reduces carrageenan-induced paw edema in rats (17) and intestinal inflammation in mice (18). CBD also counters psychoactivity, sedation, and tachycardia induced by THC (19).

SYNTHETIC CANNABINOIDS

Synthetic cannabinoids are being developed in an effort to separate psychoactivity from their analgesic and anti-inflammatory actions. The dimethylheptyl-11-oic-acid analog of CBD (DMH-CBD-11-oic acid; Fig. 1) reduces joint inflammation and tissue injury (cartilage degradation and bone erosion) in collagen-induced arthritis in mice (20). Hydrogenation of DMH-CBD-11-oic acid yields 4 distinct epimers (21). Hydrogenation at different double bonds leads to compounds with different bioactivities, none of which depend on CB1 activation. Thus, several potential therapeutic agents devoid of psychotropic activity may eventually be derived from this one phytocannabinoid. Three cannabinoids—Cesamet (nabilone; Meda Pharmaceuticals, Somerset, NJ, USA), Marinol (dronabinol; THC; AbbVie, Inc., North Chicago, IL, USA), and Sativex (*Cannabis* extract; GW Pharmaceuticals, Cambridge, United Kingdom)—which activate both the CB1 and -2 receptors have been approved by the U.S. Food and Drug Administration (FDA) for clinical use.

Nabilone (Fig.1), a dimethylheptyl analog of THC, is approved in many countries, including the United States, for treatment of the severe nausea and vomiting associated with chemotherapy. It is also used for the management of neuropathic pain and pain associated with cancer and fibromyalgia (22). It is used most commonly as an adjunctive therapy and as such results in small but significant reductions in pain.

Dronabinol, THC, was approved by the FDA in 1985 for treatment of nausea and vomiting in patients receiving cancer chemotherapy who failed to respond to conventional antiemetics. It is administered in capsule form. Dronabinol has also been used as an appetite stimulant for patients with wasting diseases such as cancer and HIV/AIDS (23).

THC with CBD (Sativex) is approved in 24 countries for the treatment of muscle spasticity associated with multiple sclerosis. Sativex was granted fast track designation by the FDA in 2014. Administered as a mint-flavored oral spray, it is now in phase 3 clinical trials in the United States for treatment of cancer-related pain. Similar to nabilone and dronabinol, Sativex treatment has the same potential adverse effects as marijuana (24). In a double-blind placebo-controlled 5 wk study of 58 patients with rheumatoid arthritis (RA) who received Sativex by oral spray (25), pain at rest, pain on movement, sleep quality, and clinical responses (disease activity score 28) were improved significantly by Sativex.

Ajulemic acid (AJA; CT-3; Resunab; Corbus Pharmaceuticals, Norwood, MA, USA) is a synthetic cannabinoid derived from a modification of THC-11-oic acid, the major metabolite of THC. Extension of the pentyl side chain from 5 to 7 carbons, addition of 2 methyl groups to increase receptor affinity, and a carboxylic acid at the 9 position to reduce blood-brain barrier penetration, results in the formation of AJA (1',1'-dimethylheptyl-THC-11-oic acid; Fig. 1 (26). AJA, administered by mouth, is 50-100 times more potent than THC as an analgesic (27). The recently developed preparation of AJA has 12 times greater affinity for CB2 than for CB1, which renders it nonpsychoactive at therapeutic doses (28). The anti-inflammatory and antifibrotic actions of AJA have been demonstrated in several in vitro systems and in animal models. Anti-inflammatory effects were first demonstrated in arachidonic acid-induced rodent paw edema (26). In an adjuvant-induced arthritis model, rats treated with 0.1 mg/kg AJA $3 \times /$ wk for 5 wk did not display evidence of active synovitis or cartilage or bone damage, whereas control animals had cartilage degradation and bone erosion that resembled RA (29). In other experiments, rats treated with AJA at up to 30 mg/kg/d for 5 d, did not show signs of physical dependence on the drug (30).

In studies designed to explore mechanisms of AJA action, it was found that addition of AJA to human peripheral blood and synovial fluid monocytes *in vitro* reduces production of the proinflammatory, bone-degrading cytokine IL-1 β (31). It is of interest that AJA did not reduce production of TNF α in these studies, given the finding that clinical trials of TNF α inhibitors in patients with systemic lupus erythematosus (SLE) have been limited by toxicity and increases in disease activity (32, 33). We have observed (unpublished data) that oral administration of single doses of 3-10 mg AJA to healthy volunteers reduced pro-IL-1 β gene expression in and secretion of IL-1 β from LPS-stimulated peripheral blood monocytes (PBMs) (Table 1; Zurier RB, Rossetti RG, Burstein SH, *et al.*, unpublished data). In contrast, AJA had no appreciable effect on TNF α mRNA levels or TNF α secretion. Maximum serum concentration of AJA reached 0.15 μ M at 5 h after a dose of 10 mg AJA (Corbus Pharmaceuticals, unpublished data).

View this table:	TABLE 1.
In this window In a new	 IL-1β secretion from stimulated PBMs from healthy volunteers administered AJA orally in
capsules	

No marijuana-like CNS effects were noted in the volunteers, as assessed by the Addiction Research Center Inventory-Marijuana (ARCI-M; developed in conjunction with the Addiction Research Center of the National Institute on Drug Abuse (National Institutes of Health, Bethesda, MD, USA) scale (34) (**Table 2**; Corbus Pharmaceuticals, unpublished data). Overall, the ARCI-M scores for treated subjects were not different from baseline scores or from placebo-treated subjects. The largest mean number of items positively responded to by subjects given AJA

http://www.fasebj.org/content/30/11/3682.full

1/9/2018

was 0.5 and occurred when the inventory was given at the second hour after administration. The largest mean number of positive responses among placebotreated subjects was 0.375 and occurred before administration. The mean response rate of placebo-treated subjects at h 2 was 0:25. These numbers are relatively small compared to those from experienced marijuana smokers. Subjects smoking marijuana report mean scores of 5.2 and, under placebo or presmoking conditions, 0.7-1.0. The relatively few positive responses obtained in this study were not dose related. These results suggest that AJA is not psychoactive at the doses tested.

View this table:	TABLE 2.
In this window In a new window	ARCI-M s
	orally in a

ARCI-M scores for AJA given orally in capsules

Addition of AJA (3-30 µM) to human monocyte-derived macrophages reduces steady-state levels of IL-6 mRNA and the subsequent secretion of IL-6 from LPSstimulated cells (35). IL-6 is a multifunctional cytokine that contributes to inflammation and tissue injury in several diseases. It has been identified in kidney biopsy tissue from patients with SLE with active glomerulonephritis (36). Skin biopsies from patients with SLE have exhibited increased expression of IL-6 in active sites (37), and plasma levels of IL-6 have correlated with lupus arthritis (38). In addition, higher levels of IL-6 in synovial fluid increase the risk of joint destruction in patients with RA (39). Because activation of osteoclasts is central to the pathogenesis of bone erosion in patients with RA, the influence of AJA on osteoclast differentiation and survival was investigated (40). Addition of AJA to cell cultures suppressed development of multinucleated osteoclasts (osteoclastogenesis), and prevented further osteoclast formation in cultures in which osteoclastogenesis had already begun. Addition of AJA to fibroblastlike synovial cells also reduces production of matrix metalloproteinases, enzymes that facilitate cartilage and bone destruction (41).

In addition to its capacity to bind to CB2, AJA binds to and activates peroxisome proliferator-activated receptor (PPAR)-y (42). PPARy receptors are members of a family of nuclear receptors that modify the transcription of target genes in response to a variety of signaling proteins. They are expressed on immune cells, such as monocytes and macrophages and regulate inflammatory responses through inhibitory effects on expression of proinflammatory cytokines and eicosanoids (43). AJA binds directly to a second site in the PPARy receptor that is separate from that utilized by other partial agonists, such as the thiazolidinediones (44). Thus, the problems of weight gain, fluid retention, and heart failure caused by the thiazolidinediones, have not been seen in animals or humans given AJA. Activation of PPARy by AJA suppresses IL-8 promoter activity. IL-8 is a chemoattractant cytokine with specificity for the neutrophil, the major cell involved in acute inflammation. Thus, suppression of neutrophil migration and reduction of enzyme release from neutrophil granules limits acute inflammation and tissue injury. The loss of PPARy in fibrotic tissues results in enhanced signaling by TGFβ, a major fibrogenic cytokine, and compounds that activate PPARy reduce fibrosis in a murine model of scleroderma (45). AJA exhibits antifibrotic effects in murine models of systemic sclerosis (46) and reduces collagen synthesis in dermal fibroblasts of patients with scleroderma (47).

The discovery of SPMs (48) has broadened our understanding of how inflammation is controlled: not simply by passive cessation of proinflammatory mediators, but also by an increase in programmed cell death (apoptosis) of immune-inflammatory cells and by activation of stop signals that lead to resolution of inflammation. It is the lack of resolution of inflammation that is in large part responsible for the signs and symptoms of diseases characterized by chronic inflammation, tissue injury, and fibrosis. Novel actions of AJA include its capacity to induce apoptosis in human T lymphocytes (49) and to increase production of 2 proresolving eicosanoids—prostaglandin (PG)J₂ and LXA₄ (Fig. 1)—that facilitate the resolution of inflammation (50, 51).

In a phase 2 proof-of-principal trial, 21 patients with neuropathic pain received twice daily doses of 20 mg AJA in a double-blind, placebo-controlled manner for 7 d (52). No clinically significant adverse events were noted. A significant reduction in neuropathic pain was noted in 30% of patients.

ENDOGENOUS CANNABINOIDS

Endocannabinoids are groups of naturally occurring members of the eicosanoid superfamily that can activate cannabinoid receptors and are derivatives of long-chain fatty acids, primarily arachidonic acid. They are produced rapidly from lipid precursors, are released from neurons by neurotransmitters or from immune cells by inflammatory agents, and can subsequently activate cannabinoid receptors on the same or on adjacent cells. Some are metabolized rapidly by the serine hydrolase FAAH to release the free fatty acid.

Anandamide (Fig. 1), the amide conjugate of arachidonic acid and ethanolamine, is one of the most important of the endocannabinoids and is well named. Ananda. from the Sanskrit word for bliss, alludes to the capacity of anandamide to increase motivation and pleasure (53). Other endocannabinoids include 2arachidonylglycerol (2-AG), and virodhamine. Enzymes known to hydrolyze endocannabinoids include FAAH, monoglyceride lipase, and N-acetylethanolamine. Endocannabinoids act to regulate inflammation and immune responses (54). Anandamide reduces mitogen-induced T- and B-lymphocyte proliferation, probably because of increased apoptosis (55). Anandamide concentrations are increased in cerebrospinal fluid and in circulating lymphocytes of patients with multiple sclerosis (56), perhaps as an attempt at regulation of the neuroinflammation characteristic of the disease. In a murine model of colitis, CB1knockout (CB1 $^{-/-}$) mice exhibited far more inflammation than animals with intact CB1 receptors (57). CB1 and -2 are up-regulated on gingival fibroblasts of patients with periodontitis. Anandamide reduces production of Porphyromonas gingivalis LPS-induced IL-6, IL-8, and monocyte chemoattractant protein-1 by these cells. Anandamide also blocks LPS-triggered activation of NFKB, a protein complex that controls transcription of DNA, cytokine production, and cell survival (58). Anandamide also suppresses TNF α -induced NF κ B activation by direct inhibition of I-κB kinase, the enzyme responsible for NFκB activation (59). Of interest is the observation that the inhibitory activity was independent of CB1 and -2 activation. Another endocannabinoid, 2-AG, appears to inhibit COX-2 via the CB1 receptor and cause down-regulation of the MAPK/NFkB signaling systems (60). Thus, further investigation and a better understanding of the regulation of endocannabinoid production and metabolism may lead to new therapy for diseases characterized by chronic inflammation and fibrosis.

LIPOAMINO ACIDS

An endogenous subfamily of eicosanoids, the lipoamino acids, are structurally and metabolically related to the endocannabinoids and also exhibit analgesic, antiinflammatory, and proinflammatory resolving properties (61). The best-studied member of this family is *N*-arachidonoyl glycine (NAgly), which is similar in structure to anandamide (Fig. 1). Indeed, oxidation of the hydroxyl group of anandamide leads to NAgly, and NAgly inhibits the FAAH-mediated metabolism of anandamide with moderate potency. There is evidence to suggest that rather than acting through the CB1 or -2 receptors, NAgly binds and activates an orphan G-protein-coupled receptor GPR18 (62). In addition, GPR18, expressed on human leukocytes, binds directly to resolvin D2 (RvD2), an immunoresolvent synthesized during the resolution phase of inflammation. In studies with mice, GPR18, bound to RvD2, stimulated macrophage phagocytosis of bacteria (*Escherichia coli* and *Staphylococcus*) and apoptosis of PMNs, thereby enhancing clearance of bacteria, limiting PMN infiltration, accelerating resolution, and reducing tissue injury. These protective actions were substantially reduced in GPR18-deficient mice (63).

It has long been thought that acute inflammation, a primitive, protective response, simply resolves—or not—on its own, spontaneously. It is now clear that just as mediators of inflammation initiate and sustain the inflammatory response, so also do lipid mediators, including LXs, resolvins, protectins, and maresins, together called SPMs, facilitate an active process of resolution of inflammation, a series of events that prevent chronic inflammation, tissue injury, and fibrosis, and promote a return of tissue to physiologic homeostasis (64). As exemplified by the animal study cited above, a deficiency of SPM impairs resolution of inflammation, just as an abundance of inflammation mediators increases the intensity of inflammation. As noted, select cannabinoids, such as AJA and NAgly, stimulate particular SPMs. In an effort to identify the precise proresolving actions of SPMs, a set of quantitative resolution indices designed to determine the active components (inflammatory and resolving) of a particular resolution process were introduced

(65). In addition, the impact of a known therapeutic agent on the resolution process can be determined. For example, down-regulation of the intracellular protein myeloid cell leukemia 1 (Mcl-1) induces apoptosis of human PMNs but does not impair their phagocytosis by macrophages, a series of actions crucial to resolution. In a murine model of bacteria-induced (*E. coli*) lung inflammation, down-regulation of inflammatory cell Mcl-1 accelerated resolution time, maintained appropriate lung function, and enhanced bacterial clearance (66). These results may in future be applied directly to treatment of patients with cystic fibrosis who experience ongoing pulmonary inflammation, even though their lung bacterial infections are cleared by antibiotics.

Just as different prostaglandins derive from different fatty acids (proinflammatory PGE₂ from arachidonic acid, anti-inflammatory PGE₁ from linoleic acid) so, too, do lipoamino acids derive from different fatty acids. *N*-linoleoyl glycine (LINgly), for example, at doses as low as 0.3 mg/kg, reduced leukocyte migration into an area of inflammation in a murine model of peritonitis (67). In addition, LINgly treatment increases production by cells in the peritoneum of the proresolving eicosanoid 15-deoxy- $\Delta^{13,14}$ -PGJ₂ (Fig. 1). A small group of *N*-linoleoyl analogs have been studied for their ability to stimulate PGJ₂ production in mouse macrophage RAW cells. The D-alanine derivative was the most active, whereas the D-phenylalanine showed almost no response. A high degree of stereo specificity was observed when comparing the D- and L-alanine isomers, the latter being the less active, a finding that suggests the response is receptor mediated.

McHugh *et al.* ($68 \Downarrow -70$) found that recruitment of BV-2 microglia by NAgly results in anti-inflammatory actions in the brain. They reported that NAgly potently acts on GPR18 to produce directed migration, cell proliferation, and perhaps other MAPK-dependent actions. These results advance our understanding of the lipidbased signaling mechanisms used in the CNS to actively recruit microglia to sites of injury. The NAgly-GPR18 pathway offers a novel approach to development of therapeutic agents to elicit a population of regenerative microglia, or alternatively, to prevent the accumulation of misdirected, proinflammatory microglia that contribute to and intensify neurodegenerative disease. These effects on microglia may also apply to inflammation in the periphery. The concept of an inflammatory reflex (71), a reflex circuit that maintains immunologic homeostasis mediated by the vagus nerve, is pertinent to a discussion of CNS regulation of inflammation. The CNS receives input from the peripheral immune system *via* inflammatory cytokines and chemokines that inform resident microglia and neurons, which in turn act to reduce further production of the cytokines. The result is that, for example, patients with RA who receive anti-TNF treatment develop changes in brain activity before resolution of inflammation (redness, swelling, heat, and pain) in the affected joints and before reduction in C-reactive protein, a circulating marker of inflammation. It appears that the nervous system is hardwired to monitor the presence of cytokines and molecular products of invaders. It may well be that the lipoamino acids promote resolution of inflammation through the inflammatory reflex. Thus, the lipoamino acids, including the cannabinoids and endocannabinoids, contain a multitude of compounds to investigate as potential new, effective, and safe treatments for diseases characterized by chronic inflammation, tissue injury, and fibrosis.

CONCLUSIONS

From the reports presented in this review, it may be concluded that several cannabinoids can be considered candidates for development as anti-inflammatory agents (Table 3). These compounds are generally free from the adverse effects associated with drugs now in clinical use. In addition, cannabinoids apparently act on inflammation through mechanisms that are different from those of other agents such as NSAIDs. A putative mechanism of action (MOA) of cannabinoids on inflammation is shown in Fig. 2, in which 2 well-studied examples, AJA and NAgly, are illustrated. The initial event is the binding to and activation of CB2 (for AJA) and GPR18 (for NAgly) at low doses in cells that are part of the immune system. In both cases an increase in release of free arachidonic acid leads to the increased production and release of proresolving eicosanoids such as PGJ₂ and LXA₄. Ultimately, this process results in an increase in the rate of resolution of chronic inflammation. These released eicosanoids may also act locally on fibroblast-like cells to reduce TGFB production and signaling, resulting in turn, in a decrease in collagen synthesis and subsequent fibrosis. At high doses, AJA can activate PPARy, which may also result in a reduction of fibrosis. As is true of all MOAs, this one will probably be modified as more data are reported. Regardless of the MOA, it

appears likely that some of the cannabinoids will be developed into safe and effective anti-inflammatory drugs.

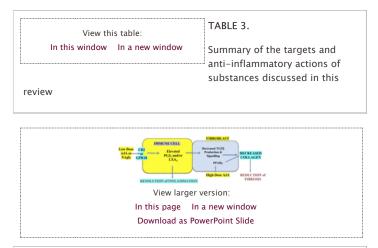


Figure 2.

A proposed mechanism for the anti-inflammatory and antifibrotic actions of selected cannabinoids is presented. Two examples are shown; the synthetic cannabinoid AJA and the endocannabinoid NAgly. The former activates the CB2 receptor, and the latter activates the orphan receptor GPR-18. In cells of the immune system, this results in increased levels of the proresolving eicosanoids PGJ₂ and LXA₄. Ultimately, this process produces an increase in the rate of resolution of chronic inflammation. A second outcome is the action on fibroblast cells, resulting in decreased collagen production and reduced fibrosis.

AUTHOR CONTRIBUTIONS

R. B. Zurier and S. H. Burstein conceived of the content of the article and wrote the manuscript.

ACKNOWLEDGMENTS

The authors thank Corbus Pharmaceuticals, Inc. for providing unpublished findings and Grant Kaufman for help in preparing Fig. 1. The authors declare no conflicts of interest.

```
2-AG
     2-arachidonoylglycerol
 AJA
مرت
ajulemic acid
ARCI-M
AKCI-M
Addiction Research Center Inventory-Marijuana
CB1/2
cannabinoid receptor 1/2
CBD
cannabidiol
COX
     cyclooxygenase
 DMH
     dimethylheptyl
FAAH
fatty acid aminohydrolase
 FDA
     U.S. Food and Drug Administration
 LINC
     N-linoleoyl glycine
 LX
     liposin
 Mcl-1
myeloid cell leukemia 1
MOA
mechanism of action
NACly
N-arachidonoyl glycine
NLINgly
NSAID, nonsteroidal anti-inflammatory drug
peripheral blood monocyte
PG
Prostaglandin
PPAR-γ
peroxisome proliferator-activated receptor-γ
RA
```

rheumatoid arthritis SLE

systemic lupus erythematosus

specializing proresolving mediators

 Δ^9 -tetrahydrocannabinol Received June 4, 2016.

Accepted July 11, 2016.

© FASEB

REFERENCES

 Busso E. P. (2007.) History of campabis and its proparations in case, science, and cobriguet *Chem. Biodivers.* 4, 1614–1648 <u>CrossRef Medline</u> <u>Google Scholar</u>

- 2. Li H -L (1975) The origin and use of campabis in Factorn Asia: their linguistic cultural implications. In *Campabis and Cylture* (Rubin, V., ed.), Mouton, The Hague, The Netherlands <u>Google Scholar</u>
- 3. Martine V (2005) Distribution of companying discontact in the control and perioderal periods custom. *Handb. Exp. Pharmacol.*, 168, 299–325 <u>CrossRef</u> <u>Medline Google Scholar</u>

4. Castanada L.T. Harvi A. Kiartechar S.M. Bath L.D. Bath M.D. (2012) Differential expression of intracellular and extracellular CP(2) cannabinoid recentor protein by human perinheral blood leukosutes. *J. Neuroimmune Pharmacol.* 8, 323–332. <u>CrossRef. Medline. Google Scholar</u>

5. Visit T. W. Newton, C. A. (2007). Therepreside potential of companying dependence drugs. Adv. Exp. Med. Biol. 601, 395-413 CrossRef Medline Google Scholar

6. Emissional and alcocanaid cignaling converses on common intracellular nathways nitric avida counting. *Prostaglandins Other Lipid Mediat.* 57, 23-34 <u>CrossRef Medline Google Scholar</u>

- 7. Sofia B. D. Kachlach L. C. Vascar H. B. (1973.) The anti-adama activity of various naturally accurring campabinoids. *Pas Commun. Chem. Pathol. Pharmacol.* 6, 909–918. <u>Medline Google Scholar</u>
- Sofia P. D. Nalaza S. D. Vassar, H. P. Knobloch, J. C. (1074). Comparative antiphlogistic activity of delta Q-tetrahydrocannahinol. hydrocortisone and achirin in various rat haw edema models. *Life Sci.* 15, 251–260 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 9. With D. W. Water F. S. Fleakly, M. Turner, C. F. Murshy, J. C. (1990) Antjinflammatory properties of compabichromene. *Life Sci.* 26, 1991–1995 <u>CrossRef Medline Google Scholar</u>
- 10. With P. W. Watcon, E. S. Elsobly, M. A. Soidel, P. Muraby, J. C. Turper, C. E. (1980). Anti-inflammatory activity of cannabichromene homologs. J. Pharm. Sci. 69, 1359–1360 <u>Medline Google Scholar</u>
- 11. Turner C. F. Flechly, M. A. (1981) Piclosical activity of compahishromone, its homology and icomore. J. Clin. Pharmacol. 21(8–9, Suppl)283S-291S CrossRef Medline Google Scholar
- Malfait A. M. Callille, B. Sumarius IIa, B. F. Malik, A. S. Andraskas, F. Machaulan, B., Foldmann, M. (2000). The nonnextchartive company: constituent companyidial is an availantic atheritic therapeutic in myrine collagen-induced arthritis. *Proc. Natl. Acad. Sci. USA* 97, 9561–9566
- Purstein S. Vasanelli C. Slada L. T. (1975.) Prostaglanding and cannabic-III: inhibition of biosynthesis by assantial ail components of marihuana. *Biochem. Pharmacol.* 24, 1053–1054. <u>CrossRef. Medline. Google Scholar</u>
- 14. Purstain S. Taulor B. El Earaly E. S. Turper C. (1076) Prostalanding and companies -V: identification of n-vinulohenol as a notent inhibitor of prostalandin synthesis. *Biochem. Pharmacol.* 25, 2003-2004 <u>CrossRef</u> <u>Medline Google Scholar</u>
- 15. George H. L. Lutzin, L.M. Solamink, C.A. Nusteren, D. H. (1072) Inhibition of prostoclandin biocumthesis hu derivatives of clivital formed under nurolysis of compahidial *Biochem. Pharmacol.* 27, 607–608 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 16. Pardan P. K. (1986) Structure activity relationships in cannabinoids. *Pharmacol. Rev.* 38, 75-149 <u>Medline</u> <u>Google Scholar</u>
- 17. Costa P. Ciannani C. Eranka C. Trovata A. E. Colleani M. (2004) Vanilloid TDDVI recentor mediates the antihyneraleasic effect of the nonpsychoactive companied compahidel in a rat model of acute inflammation. *Br. J. Pharmacol.* 143, 247-250 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>

Priste M. Multichendhum, D. Périst C. Harlif, C. Harliff, C. Harl

Heart Circ. Physiol. 293, H610-H619

- 19. Dura E. C.W. (2006) A tale of two compliancies the thermostic rationale for combining totrabudrocannabinel and compliancies. *Med. Hypotheses* 66, 234-246 <u>CrossRef Medline Google Scholar</u>
- 20. Superinglia B.E. Callik B. Tchiliban S. Erida E. Machaulam B. Ealdmann M. (2004.) A noval synthetic nonnexicharctive cannabinaid acid (HIL-220) with antiinflammatory properties in murine collagon-induced arthritis. *Arthritis Rheum.* 50, 985–998. <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 21. Den Shahet S. Hanus J. O. Katanian G. Callib, B. (2006) New comparidad derivatives: curthesis, binding to comparing directory and evaluation of their antiinflammatory activity. *J. Med. Chem.* 49, 1113–1117 <u>CrossRef</u> <u>Medline Google Scholar</u>
- 22. Teaco C. C. Ciudice M. C. (2016) Nabilana for the management of pain. *Pharmacotherapy* 36, 273-286 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 23. Podowski, M. E., Boroz, S. E. (2016.) Clinical utility of dropphinal in the treatment of weight loss associated with HIV and AIDS. *HIV AIDS (Auckl.)* 8, 37-45 <u>Medline Google Scholar</u>
- 24. Duran M. Nara A. Jon A. Sorra F. D'Alan C. Bramanti B. Calabid, P. S. (2016) Evaluating Sativax[®] in neuropathic pain management: a clinical and neurophysiological assessment in multiple sclerosis. *Pain Med.* 17, 1145-1154 <u>Medline Google Scholar</u>
- 25. Plaka D. P. Bahan D. Ha M. Juhh D. W. Macaha C. S. (2006). Braliminary madicine (Sativari) in the treatment of pain caused by rheumatoid arthritis. *Rheumatology (Oxford)* 45, 50-52
- Purstain S. H. Audatta C. A. Brauar A. Davana W. A. Caladnar S. Davla S. A. Mashaulam B. (1992.) Sunthatic nonpsychotronic cannahinoids with notant antiinflammatory analysis and laukasute antiadhesion activities. J. Med. Chem. 35, 3135-3141 CrossRef Medline Google Scholar
- 27. Purstein S. (2005) Aiulemic acid (IP-751): sunthesis, proof of principle toxicity studies, and clinical trials. *AAPS J.* 7, E143-E148 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 28. Terrer M. A. Zurier, P. P. Durther C. H. (2014) Ultranura similaria acid has improved CP2 coloctivity with raduced CB1 activity. *Bioorg. Med. Chem.* 22, 3245-3251 <u>CrossRef Google Scholar</u>
- 29. Turior P. P. Borcotti, P. C. Lano, L. H. Coldborg, L. M. Hunter, S. A. Purstein, S. H. (1998) Dimethylantyl_THC-11 oic acid: a nonneyschoactive antiinflammatory acent with a cannabinoid template structure. *Arthritis Rheum.* 41, 163-170 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 30. Deter K. S. Deter K. B. Teder J. Deter N. C. Schwart T. C. Nedera G. D., Tarley M. S. Deter M. T. Mir C. N. (1000.) 17 17 Dimethylkoptyl dolta 8 totrahydrocompahinal, 11 aic acid: a noval arally affactive compahinaid with analogs: and anti-inflammatory properties. *J. Pharmacol. Exp. Ther.* 291, 31-38
- 31. Zurier B. P. Berretti B. C. Burretain S. H. Bidinear B. (2002) Suppression of human monositie interfaultin. These production by sindemic acid a nonneuropartitie connabinoid. *Biochem. Pharmacol.* 65, 649-655 <u>CrossRef</u> <u>Medline Google Scholar</u>
- 32. Aringer M. Houssiau E. Cordon C. Craninger W. P. Voll P. E. Path E. Steiner G., Smalan J. S. (2000). Adverse events and afficacy of TNE-alpha blockade with inflivimab in patients with systemic lunus anthematosus: long-term followup of 13 patients. *Rheumatology (Oxford)* 48, 1451-1454
- 33. Welliams E. L. Cadala G. Edwards C. L. (2009) Anti-TNF-induced lupus. *Rheumatology (Oxford)* 48, 716-720
- Harden G. A. (1065.) Addiction Becarch Conter Inventors (ABCI): development of a conterl drug actimation scale. J. Nerv. Ment. Dis. 141, 300– 307 CrossRef Medline Google Scholar
- 35. Barker L. Ataz E. Bossatti B. C. Skulas A. Batel B. Zurier B. P. (2008) Suppression of human macrophage interleukin-6 by a nonposychoactive campabinoid acid. *Rheumatol. Int.* 28, 631–635 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 36. ******* Cmolen, J. S. (2005.) Cytokine expression in lupus kidneys. *Lupus* 14, 13-18
- 37. Mikita N. Jkoda T. Johanna M. Europhysics E (2011) Pacent advances in establines in cutaneous and systemic lupus erythematosus. *J. Dermatol.* 38, 839–849 <u>Medline</u> <u>Google Scholar</u>
- 38. Pall E. M. Cibcon, D. S. Pall A. L. Boonau, M. P. (2014). Plasma II –6 levels correlate with clinical and ultracound measures of arthritis in patients with systemic lupus erythematosus. *Lupus* 23, 46–56
- 39. Kotoko C. Soto K. Kim K. L. Tokohotki N. Udogowa N. Nakamura L. Souther and K. Kim K. L. Tokohotki N. Udogowa N. Nakamura L. soluble interleukin-6 receptors in the synovial fluids from rheumatoid

Min. Res. 11, 88-95 CrossRef Medline Google Scholar
40. Concert K. L. Soltman, L. L. Stein, C. S. Lian, L. B. Zurian, B. B. (2008) Adulantic and induced and induced another in the concertainty of the scholar scho

arthritis patients are responsible for osteoclast-like cell formation. J. Bone

- Johnson D. P. Stabulis L.A. Bossetti, P. C. Burstoin S.H. Zuriar, P. B. (2007)
 Suppression of fibroblast metalloproteinases by aiulemic acid. a nonsychoactive campabinoid acid. *J. Cell. Biochem.* 100, 184-190 <u>CrossRef</u> Medline Google Scholar
- 42. Lin L. Li L. Burstein S. L. Zurier, P. P. Chan, L. D. (2002.) Activation and binding of neroviceme preliferator, activated recenter common by synthetic cannabinoid ajulemic acid. *Mol. Pharmacol.* 63, 983–992
- 43. O'Sulliver C. F. Kardell, D. A. (2010.) Connection of activation of inflammatory disease. *Immunobiology* 215, 611-616 <u>CrossRef Medline</u> <u>Google Scholar</u>
- 44. Ambracia A L. Dias S. M. Balikarnov, L. Zuriar, P. P. Burstain, S. H. Carratt, P. C. (2007.) Aiulemic acid: a sunthatic nonnexycloactive cannabinoid acid, hound to the ligand binding domain of the human perovisione proliferator-activated receptor gamma. *J. Biol. Chem.* 282, 18625-18633.
- 45. Will M. Molichian, D. S. Chang, E. Warner, Plankanshin, M. Chash, A. K. Marga, L. (2000). Postialitazona abrogatas blaomycin-induced sclarodarma and blacks. profibratic responses through perovisione proliferator-activated recentorgamma. Am. J. Pathol. 174, 519-533 CrossRef Medline Google Scholar
- 46. Lucattalli M. Einstein C. Solui F. Carrie Consolat F. Pattalati P. De Curta C., Larantini S. Calanti M. Lucatalla C. (2016) Audomic acid events notant anti-fibratic effect during the fibration phase of bleomycin lung. *Respir. Res.* 17, 49 <u>CrossRef Medline</u> <u>Google Scholar</u>
- 47. Conzolaz E C. Solvi E. Policitori E. Alematchina A. Bolumbo K. Loronzini S. Lozzarini B. E. Montilli C. Conoschi B. L. Lucattalli M. Boldi C. Cionchaschi E., Colonzai M. Bocini E. L. Distlar L. H. (2012.) Synthetic connobinoid aiulemic acid everts notent antifibratic affects in experimental models of systemic sclerosis. Ann. Rheum. Dis. 71, 1545–1551
- Bucklass C. D. Cilcass D. W. Sarbar, C. N. (2014). Bronarching linid mediators and machanisms in the resolution of acute inflammation. *Immunity* 40, 315– 327 <u>CrossRef Medline Google Scholar</u>
- Pidiana B. Tarres B. B. (2002) Aiulomic acid a popportehoactive composition of a source o
- 50. Zurier P. P. Sun Y. P. Coorea, K. L. Stabulis, L.A. Bossatti, P. C. Skulas, A. Judas, E. Sarban, C. N. (2009.) Aiulemic acid, a sunthatic cannabinoid, increases, formation of the endogenous prorecolving and anti-inflammatory eicosanoid, lipoxin A4. FASEB J. 23, 1503–1509.
- 51. Stabulis L.A. Johnson, D.B. Bossatti, B.C. Burstein, S. H. Zuriar, B.B. (2008) Audamic asid a sumthatic compahinaid asid induces on antiinflamm atory arcfile of siscenseide in human surrovial cells. *Life Sci.* 83, 666-670 <u>CrossRef Medline Google Scholar</u>
- 52. Veret M. Celler K. Burthelin C. Connel L. Have C. Cherider H. (2002) Analgesic offact of the cumbatic connection of CT. 2 on chronic neuropathic pain: 2 randomized controlled trial. *JAMA* 290, 1757–1762 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 53. Montoleone A. M. Di Marzo V. Austa T. Dissitelli, E. Dalla Craue P. Scannamialia B. El Chech M. Caluai S. Montoleone P. Mai M. (2015). Deranged andocannabinoid reconnects to bedonic opting in underweight and recently. weight-restored patients with anorexia nervosa. Am. J. Clin. Nutr. 101, 262– 269
- 54. Shaper S. D. Di Marze V. (2012) Endocennabinoids in norvous system health and diseases the big nicture in a nutshell. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 367, 3193-3200
- apoptosis. J. Neuroimmunol. 55, 107-115 CrossRef Medline Google Scholar
- 56. Contonzo D. Pari M. Possi S. Brosnorstti C. Eurlan P. Enza E. Do Chiara V., Pattistini I. Pornardi C. Pornardini S. Martino C. Massarrono M. (2007.) The endocompabinoid system is dysrogulated in multiple sclerosis and in experimental autoimmune encephalomyelitis. *Brain* 130, 2543-2553
- 57. Marco C. Marco M. Line P. (2004) The endocronous competingid custom protocts acounts colonis inflammation. *J. Clin. Invest.* 113, 1202–1209 <u>CrossRef Medline Google Scholar</u>
- 58. Nakaiima V. Eusuichi V. Biswas K. K. Hashiaushi T. Kawahara K. Vamaii K. Uchimura, T., Izumi, Y., Maruyama, I. (2006) Endocannabinoid, anandamide in

- cincipal tissue regulates the periodental inflammation through NF-kappaB pathway inhibition. *FEBS Lett.* 580, 613-619 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 59. Carte B. Calada M. A. Di Mara V. Anardia. C. Muita F. (2002) Anardamida inhibita nuclear factor konnell activation through a cannabinoid receptor-independent pathway. *Mol. Pharmacol.* 63, 429-438
- 60. Zhang J., Chan C. (2008.) Endocannahingid 2-arachidonovlalveeral protects neurons by limiting COX-2 elevation. *J. Biol. Chem.* 283, 22601-22611
- 61. Burstein S. H. (2014). The companying of a cide analogs and and account counterparts. *Bioorg. Med. Chem.* 22, 2830–2843. CrossRef Google Scholar
 62. Burstein S. H. Macuria, C. A. Pare, A. H. Calmanus, P. A. Zuria, P. G. (2011).
- Becolution of inflammation by N. arachidanou/glycine. J. Cell. Biochem. 112, 3227-3233 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 63. China N. Dalli J. Colar B. A. Sorban C. N. (2015.) Identification of resolution D2 recenter mediating resolution of infections and organ protection. *J. Exp. Med.* 212, 1203–1217
- 64. Serban C. N. (2014) Pro-resolving lipid mediators are leads for resolution physiology. *Nature* 510, 92-101 <u>CrossRef Medline</u> <u>Google Scholar</u>
- 65. Schurch L. M. Chinas N. Avita M. Sarban C. N. (2007) Basalvin E1 and protectin

<u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>

66. Lucas C. D. Donward, D. A. Tait, M. A. Eox, S. Manuick, J. A. Allon, K. C. Bobb, C. T. Histori, N. Haslatt, C. Duffin, P. Bossi, A. C. (2014). Downroadulation of Mel-1 has anti-inflammatory pro-resolution effects and enhances bacterial clearance from the lung. *Mucosal Immunol.* 7, 857–868 <u>Medline</u> <u>Google Scholar</u>

- Burstein S. McOusin C. Salmanson P. Saical P. (2012) N-Amina acid linelaov(conjugatos: anti-inflammatory activities. *Bioorg. Med. Chem. Lett.* 22, 872– 875 <u>CrossRef Medline Google Scholar</u>
- Mattheb D. Hu, G. C. Bismannen, N. Julian, A. Vanal, Z. Wallow, M. B. Bradahaw, H. B. (2010). N. arashidanovil alusina, an abundant and asanovis lipid, advantable drives directed callular microtion through CBP19, the nutative abundant completion in the second callular microtion through CBP19, the nutative abundant completion in the second callular microtion through CBP19, the nutative abundant completion in the second callular microtion through CBP19, the nutative abundant completion in the second callular microtion through CBP19, the nutative abundant completion in the second callular microtion through CBP19, the nutative abundant completion in the second callular microtion in the second callular microtion in the second callular abundant completion in the second callular microtion in the second callu
- 69. McHuch D. Wasser Miller L. Passe L. Prodebaw, H. B. (2012) siPNA knockdown of CDP18 recentors in PV-2 microalia attenuates N-arachidonovi glycineinduced cell migration. J. Mol. Signal. 7, 10 <u>CrossRef</u> <u>Medline</u> <u>Google Scholar</u>
- 70. Mellich D. Base I. Dune E. Bredehnin H. B. (2012) A(0). Totrahidrocannabinol migration in human andomatrial HEC. 19 cells. *Br. J. Pharmacol.* 165, 2414-2424 <u>CrossRef Medline Google Scholar</u>
- 71. Diemend B. Terem K. J. (2011) Manning the immunological homunculus. Proc. Natl. Acad. Sci. USA 108, 3461–3462

We recommend

Cannabidiol, unlike synthetic cannabinoids, triggers activation of RBL-2H3 mast cells. Elda Del Giudice et al., J Leukoc Biol

Mechanisms of Tolerance to delta-9-THC in Rodent Models of Pathological Pain

Daniel Morgan1 et al., FASEB J

Fish-oils Increase BAMBI Expression to Protect Against Fibrotic Activity in LPS Stimulated Hepatic Tissue Megan L. Schaller et al., FASEB J

Sex differences in cannabinoid antinociception against inflammatory pain

Rebecca M Craft and Ram Kandasamy, FASEB J

The Non-psychoactive Phytocannabinoid, Cannabidiol (CBD), and the Synthetic Derivatives, HU308 and CBD-DMH, Reduces Hyperalgesia Natural cannabinoids from omega-3 fatty acids combat inflammation Catharine Paddock PhD, Medical News Today

Cannabinoids and cancer: causation, remediation, and palliation Wayne Hall et al., The Lancet Oncology

Marijuana (Cannabis): Facts, Effects and Hazards Kathleen Davis FNP et al., Medical

News Today

NSAIDs (Non-Steroidal Anti-Inflammatory Drugs) MyVMC

Turned-Off cannabinoid receptor turns on colorectal tumour growth MvVMC

and Inflammation in a Mouse Model of Corneal injury. Dinesh Thapa et al., FASEB J

Powered by